

# Statistical Mechanics of Horizontal Gene Transfer in Evolutionary Ecology

Nicholas Chia · Nigel Goldenfeld

Received: 29 October 2010 / Accepted: 13 December 2010 / Published online: 4 January 2011  
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**Abstract** The biological world, especially its majority microbial component, is strongly interacting and may be dominated by collective effects. In this review, we provide a brief introduction for statistical physicists of the way in which living cells communicate genetically through transferred genes, as well as the ways in which they can reorganize their genomes in response to environmental pressure. We discuss how genome evolution can be thought of as related to the physical phenomenon of annealing, and describe the sense in which genomes can be said to exhibit an analogue of information entropy. As a direct application of these ideas, we analyze the variation with ocean depth of transposons in marine microbial genomes, predicting trends that are consistent with recent observations using metagenomic surveys.

**Keywords** Evolution · Horizontal gene transfer · Mobile genetic elements · Metagenomics

## 1 Introduction

The advent of high throughput sequencing technologies and their application to genomics and metagenomics provides an ever-growing torrent of data that is providing fine-grained data about ecosystems and is beginning to alter our views on biological evolution [21, 36]. In particular, it is becoming clear that the biological world, especially its majority microbial component, is strongly interacting and may be dominated by collective effects. Such phenomena can arise of course through cell-to-cell communication by signaling molecules [89], but there is also a genetic mode of communication: genes can be transferred between living

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N. Chia (✉)  
Institute for Genomic Biology, University of Illinois at Urbana-Champaign, 1206 West Gregory Drive,  
Urbana, IL 61801, USA  
e-mail: [chian@uiuc.edu](mailto:chian@uiuc.edu)

N. Goldenfeld  
Loomis Laboratory of Physics, Department of Physics and Institute for Genomic Biology, University of  
Illinois at Urbana-Champaign, 1110 West Green Street, Urbana, IL 61801, USA  
e-mail: [nigel@uiuc.edu](mailto:nigel@uiuc.edu)

cells not related by heredity, and subsequently expressed. Comparative genomics indicate the widespread and frequent presence of this so-called “horizontal gene transfer” (HGT) between organisms, not only those that are closely-related but also those that are distant taxa. Metagenomic surveys also quantify the unsuspected extent of the abundance of mobile genetic elements (MGEs) such as plasmids, viruses, and transposons, to a degree that exceeds organismal abundances by an order of magnitude in many different ecosystems [24, 29].

The basis of classic Neo-darwinist population genetics rests on the notion of gradual differences arising (i.e., via point mutation) within a population. Populations are defined by firm geological and species barriers that constrain the flow of genetic information. By contrast, mobile genetic elements (MGEs) such as plasmids [76], viruses [67], and transposons [57] are capable of producing large genomic changes and can jump between (classically defined) populations by crossing species barriers. In particular, horizontal gene transfer—the transfer of genes between organisms that are not related by heredity—creates novel gene combinations by shuffling genetic material between organisms that share the same environment. Such gene transfers can have an enormous impact on the process of biological evolution [4, 5, 29, 32, 75, 81].

The growing realization of the widespread nature of horizontal gene transfer (HGT) is bringing about a re-evaluation of some of the fundamental notions in biology, such as the species concept [33, 34]. Drawing general conclusions about the frequency and effects of HGT requires massive amounts of genome data. Experimentally, advances in high-throughput sequencing have allowed researchers to sequence environmental samples identifying microbial taxa and gene compositions [51].

The lessons from statistical physics can be fruitfully translated to evolutionary biology. Methods from statistical physics often utilize an abundance of data in order to draw out general characteristics and conclusions about the central features that describe system behavior. Statistical approaches have already been implemented for many biological systems including metabolic networks [46, 83] and ecology [23, 43]. Statistical physics is potentially important for understanding the emergent properties of dynamical biological systems, such as the characteristics and key features of evolution with HGT. Collective phenomena such as synchronization in fireflies [61] or pattern formation in predator-prey dynamics [13] rely on techniques for studying many individual trajectories in order to arrive at a condensed, more principled, understanding of system behavior. HGT can be thought of as an interaction that couples the many potentially different genomes in a population of organisms. Indeed, cellular evolution as a whole finds the language of statistical physics to often be the most appropriate for describing how modern cells evolved [93, 94].

In the remainder of this article, we briefly introduce the biology of horizontal gene transfer and describe some of the ways in which HGT plays an important role in biological systems. We then describe how HGT requires an extension of “classical” notions of evolution, and how this is being accomplished by recent efforts to model the effect of HGT on genetic evolution. In order to predict the outcomes of HGT, concepts from statistical physics can be useful, for example in interpreting ecological data. In particular we consider a striking trend that has emerged from the sampling of environmental DNA extracted from marine microbes, namely the variation in density of a particular type of mobile genetic element (transposons) with depth in the ocean. We argue that an intuitive notion of genome entropy, describing the variable regions of a genome (i.e. not the core, conserved genes), leads to trends very similar to those observed.

## 2 The Extent of Horizontal Gene Transfer

Horizontal gene transfer, or HGT, is a ubiquitous feature of genome evolution [25]. For the purposes of this article, we consider an event to be a HGT if it involves any transfer or introduction of any genetic material that does not stem from cellular replication. Evidence for HGT is wide-ranging. It occurs within and between all domains of life [31, 42, 49, 62, 64]. The range of HGT encompasses the entire scale of organismal complexity, from viruses [40] to multicellular eukaryotes [49]. Time is apparently not a barrier: HGT events ranging from ancient [96] to very recent [41] have been reported. Indeed, there appears to be no absolute barrier to HGT, and we conclude that it is a generic feature of genome dynamics.

Microbial genomes also contain and interact with a large variety of mobile genetic elements (MGEs) including plasmids [76], viruses [67], and transposons [57]. Counts of horizontally transferred genes identified from G+C content found them to make up anywhere from 1.5 to 14.5% of microbial genomes [30]. Characterizations of HGT in different gene families found that 34% of all gene families were identified as having undergone HGT at some point in their evolutionary history [18].

Examples of genes transferred between evolutionarily distinct microbes include rhodopsins in marine bacteria and archaea [20, 28]. These photosynthesis genes have been linked not to a particular organism so much as a particular environment—so-called ‘cosmopolitan genes’ [28]. These genes appear to be readily transferred and incorporated into many different microbes, and appears to aid them in harvesting light. The idea of a gene being adapted and more tenaciously linked to an environment than a particular microbe departs radically from the more classical notions of vertical descent with mutation. In such cases, we are taking implicitly a more gene-centric than organism-centric view of evolution.

While examples such as the photosynthetic rhodopsins depict a fairly harmless tale of environmentally adapted genes, the same evolutionary mechanisms are also responsible for a medical disaster that has arisen due to the lack of appreciation for the potential swiftness of evolution. Antibiotic and drug resistance genes rapidly adapt to new hosts, meaning that once a single population of pathogens becomes resistant to treatment, potentially many more rapidly will acquire resistance soon thereafter [10, 70]. Moreover, once the genes for antibiotic or drug resistance develop in some pathogen they seem to persist over long periods of time in some form. Thus, while avoiding a particular antibiotic treatment for long periods of time has resulted in the loss of resistance, very rapid re-emergence of resistance occurs once the antibiotic is reintroduced [70]. Interestingly, HGT of antibiotic resistance genes from antibiotic-producing bacteria does not appear to have played the major role in the evolution of antibiotic resistance in the clinical setting [3]. Instead, most antibiotic resistance genes appear to have originated and diversified in other environmental bacteria. They were then disseminated widely, and these underlying genes formed the basis for the development of antibiotic resistance in pathogens and commensal bacteria [3]. Resistance that seems to have been developed in commensal microbes has made its way back to more open environments in soil and water [50].

Potential barriers to HGT appear to be abundant [82]. First, genes must be delivered into a microbe. This already presents a number of barriers. Viruses and plasmids have limited host ranges [26]. Naturally competent microbes such as *Neisseria* are capable of uptaking raw DNA from their environment (a process known as transformation), but uptake is limited to DNA containing certain sequence motifs [14, 55]. Genes which are not favorable are rapidly deleted and lost from a genome [74]. In order to be retained on longer timescales, acquired genes must pass through a gamut of hurdles. In order to benefit its host, the newly-introduced gene must somehow be expressed. Depending on the gene’s history, it may now

be subject to a new regulatory scheme. Even once expressed, the gene may not be expressed with the right timing or in the right amounts to benefit its new host. Moreover, the newly expressed product has not yet been adapted to the host environment, which may lead to a greater chance of unwanted or deleterious interactions with other proteins in the new cellular environment. Ultimately, the horizontally-transferred gene must bypass the many levels of regulation and positively impact the organism it has entered in a short time in order to be retained.

These barriers appear to be more surmountable the more closely related are the donor and recipient, and as such, HGT is generally believed to occur more often between the same or similar species [26]. This tendency is amplified by the fact that organisms of the same species are spatially-correlated also. However, overall there appear to be no absolute barriers to HGT despite the numerous barriers to any individual trial. Along with how many HGT events are retained in the long term, another pertinent question to raise is how frequently does HGT occur on a shorter timescale? By directly counting the number of times each plasmid horizontally transferred in a population of lab-grown *Escherichia coli*, Babic et al. were able to determine that plasmids were transferred approximately once per cell generation [6]. Thus, while the barriers to individual HGT events persisting in a foreign host for very long might be low, the number of attempts are seemingly high.

### 3 Detecting Horizontal Gene Transfer

Despite the extent of HGT, the evolutionary history of organismal lineages is preserved through the consistent phylogenies derived from a number of core subsystems including translation, transcription, and DNA replication [12, 17, 95]. These subsystems are highly-conserved and present in every cell. The consistent pattern of relationships within these major cellular subsystems defines microbial taxonomy. HGT can then be detected as gene relationships that differ greatly from this canonical organismal phylogeny. Horizontal movement of genetic material is detected by comparing the evolutionary relationships between genes in different species against their organismal relationships. This can be done using a number of metrics including correlations in sequence distance [27], phylogeny [7, 53], or gene composition [19, 84].

It is noteworthy that many of these methods rely on properly characterizing the statistical distributions of gene properties [19, 27, 84]. HGT detection methods generally rely on there being significant differences in organismal and gene pattern of descent. Limitations on resolution within the organismal phylogeny, or evolutionary relationships, makes it difficult to detect HGT between closely related species. Thus, HGT is nearly undetectable between the organisms for which it is expected to occur the most frequently. Since we lack the resolution to distinguish organismal relationships between closely related species, we cannot track their HGTs through sequence analysis. Theoretical models of the role of HGT in biology becomes all the more important for discerning both the extent and possible effects of HGT on organismal evolution.

What sequence analysis cannot see in nature, fluorescence microscopy has managed to visualize in the laboratory. By tracking an *Escherichia coli* plasmid with a fluorescent marker, Babic et al. were able to directly visualize HGT [6]. While these studies do not tell us much about the general rate of HGT between species, they can measure the level of activity of particular MGEs, providing us with a general idea of what the limits of HGT might be.

HGT may also be detectable indirectly through its influence on genome organization. Genomes are not unstructured chains of genes, but apparently possess an architecture that,

particularly in the case of microbes, can assist in gene expression and genome evolution. One of the most frequently-encountered structures in biology is modularity: a complex network (e.g. metabolic or gene regulatory) that can be decomposed into independent (i.e. weakly-interacting) internally-connected functional parts that can evolve separately with minimal disruption to the system as a whole [37, 52, 73, 88]. Modular networks can arise when the environment is fluctuating in time, creating a modularly-varying potential for the dynamics [47], or more generally, selecting for the organizational structure that can change in the most facile manner [66]. For a similar reason, a spatially-heterogeneous environment, such as might arise after an extinction event, can also promote the emergence of modularity [48] as a suddenly vacated ecological niche becomes available for colonization. One way for modularity to arise is through the horizontal transfer of a gene or collection of nearby genes that code for a particular part of a network [38, 56, 78], thereby accelerating the dynamics. There is evidence that networks can indeed grow by acquiring genes in groups (known as operons, known to govern coupled reactions in the cell), and that these are attached preferentially at the edges of the existing network [65].

#### 4 Modeling Horizontal Gene Transfer in Biology Using Statistical Mechanics

MGEs such as viruses outnumber microbes by over 10-to-1 in environmental samples [79, 90]—a fact suggestive of their larger role in microbial ecology [80] and evolution [29, 32]. Assessing the role of these MGEs in the ecosystem, however, is difficult. Doing so by direct experimental measurements, such as sequencing, is essentially impossible. Even if one knew every environmental DNA sequence and biochemical reaction within an environment across time, the task of reconstructing the effect of HGT on the entire ecosystem would be akin to calculating the structural soundness of a skyscraper on the basis of the positions and properties of its elemental particles.

The importance of studying a problem at an appropriate scale applies to biology as well as physics. The goal of ecology is to understand the principles underlying change and stability of populations. For example, the role of a microbial species is something of consequence, whereas the role of an individual microbe is not. Ecological modeling seeks to answer questions about the general nature of local processes that give rise to global behaviors or properties. Understanding how local and global processes are related can give us an outline of the forces driving the system.

There are a number of examples in the literature of insight gained from coarse-graining biological systems, studied using statistical mechanics ideas ranging from spin glass models [38, 78] to non-equilibrium statistical mechanics. An example of the latter is the phenomenon of speciation. For example, analysis of the codon usage of genes extracted from libraries of *Escherichia coli* strains indicate that speciation may have arisen as a result of HGT [59]. A statistical mechanical model shows specifically how HGT can give rise to speciation—global genome sequence divergence—in a population of closely-related organisms by seeding the propagation of mutational fronts [86]. In this study, statistical mechanics was used to model the system-wide consequences of the interplay between point mutation and homologous recombination, following a single HGT event in microbes. In asexual organisms, such as bacteria, homologous recombination allows genome sequences to be repaired and thus made more uniform in a population, while point mutations are a source of genome disorder. Species-specific biological details are important, because the successful insertion of a piece of alien DNA in a pre-existing bacterial genome relies on the ends of the insertion matching with both surrounding pre-existing base pairs. This requirement of

matching ends is absent in some bacteria, or only enforced at one end in others. In addition, the cell has mechanisms to prevent the insertion of alien DNA, but these mechanisms become less effective the closer the alien DNA sequence matches the region it replaces. The behavior of such a population of interacting genomes can be explored by Monte Carlo simulation, and as a function of the rates of point mutation and recombination, the phase diagram mapped out. Interestingly there is a generically first-order phase transition between a state with a monodisperse population and a state with a diverse population. The transition occurs through the propagation of what have come to be known as diversification fronts propagating along the genome over the course of evolutionary time. The front propagation arises because around an insertion, the disruption of the canonical sequence means that recombination is locally suppressed, leading to the build-up of point mutations and the extension of the region of sequence divergence. Such fronts are predicted to occur in strains of *Bacillus cereus* but not in strains of *Buchnera aphidicola*, owing to the details of their mismatch repair mechanisms, and these predictions were confirmed by comparative genomics studies on the fully-sequenced genomes of these organisms. This mechanism for speciation would leave behind a genome that has a mosaic structure, corresponding to the merging of several diversification fronts arising from distinct horizontal gene transfer events. Indeed, such puzzling genome features have been observed in an environment where naively one would have expected extreme selection to have provided essentially no diversity [85]. The mechanism discussed here is especially interesting, being a counterexample to the popular notion that speciation arises purely as a result of Darwinian selection.

Other features have been linked to HGT through modeling, including modularity [78], the optimality of the genetic code [87], and phase variation of biofilms [16] (a microbial analogue of multicellular differentiation). Population and gene heterogeneity within an environment is counterbalanced in microbial systems by mechanisms such as homologous recombination that serve as an additional homogenizing force [92].

HGT impacts the safety of the biotechnology industry greatly. Monitoring and modeling the spread of genes such as virulence factors enhances public safety and helps the development of better lab practices [63]. In addition, MGEs can have impact beyond the movement of genes between organisms. In oceanic carbon cycling, viruses are thought to “kill the winner” since having the most available hosts should positively feedback into greater predation [80]. This dynamic may play an important role in the diversification of ocean microbes by removing dominant species [91] and in the boom-bust cycles of planktonic blooms, some of which can extend for hundreds of square miles [72]. Both questions are important to biodiversity and ecological stability. Plankton blooms choke off oxygen and nutrients within a large portion of the ocean and may be the result of trace minerals or other man-made pollutants [9, 35]. The factors regulating these boom-bust cycles are a topic of current debate [8, 44, 60].

## 5 Horizontal Gene Transfer and Genome Entropy

Quantitative modeling shows that the early benefit from HGT can explain certain general properties of biology including the emergence of a universal genetic code [87] and modularity [78]. Modern life involves a complex web of enzymatic interactions bringing additional interaction and regulatory barriers to HGT. However, examples of recent beneficial HGT are abundant and reveal that HGT is still occurring in modern organisms. HGT serves as an effective means for modulating mutation rate within an organism. Modeling shows that mutation rate is indeed selectable [22]. Examples of organisms altering their mutation rates

include the SOS response in *Esherichia coli* [58] and the competence response of *Bacillus subtilis* [39].

HGT accelerates organismal evolution by allowing for the exchange of genes between two organisms, populations, or species. HGT modifies the types of genetic changes and enhances the amount of total genetic mutation. In that sense, HGT can be thought of as a source of disorder in a genome, in effect raising its information entropy [1]; thus, we will informally use the notion of a genome “temperature” to represent the level of disorder in a genome. Analogies between HGT and temperature have been made in the context of studies of the evolution of biological complexity using digital organisms [2], and also in the evolution of cells and genomic annealing [93]. Genomic entropy roughly correlates to the amount of genomic change per unit time. This directly affects both the rate of information loss and organismal adaptation. Genomic annealing refers to the concept that early forms of life had few barriers and much to gain from HGT. This resulted in massive HGT that slowed down or became quenched as barriers arose due in part to the increasing complexity of cellular life.

At first glance, these two aspects of genome “temperature” appear to be quite different. However, they can be understood as different sides of the same coin. Genomic entropy is a property that reflects environmental information. For success, changes to the environment must be met with organismal adaptation. In other words, the nature of genome plasticity must be reflective of fluctuations in the environment. At the same time, not all aspects of the environment are in constant flux. Ideally a genome would keep well-adapted genomic elements constant and only change those that need to change. This is the concept that leads to genomic annealing, whereby barriers to entropy (in the form of HGT) arise from increasing complexity. Here we will not attempt to address issues of how to define complexity; instead, we will attempt to see if the concept of genome entropy in the information sense can be helpful phenomenologically.

Competition and changing environments dictate that organisms that can quickly adapt to these ever changing circumstances will hold an advantage over their neighbors. It seems natural that a readily available adaptive mechanism such as HGT would be utilized for exactly these reasons.

While the frequency of HGT in environmental microbes is inaccessible through direct measures, the importance of maintaining evolutionary “temperature” can be inferred from metagenomic surveys of the ocean [51]. Kostantinidis et al. sequenced over 200 Mbp of a random whole-genome shotgun (WGS) library obtained from a depth of 4,000 m at Station ALOHA in the Pacific Ocean. The per-bp density of a typical type of transposon known as an insertion sequence (IS), which can be identified by genes that codes for Transposase, was measured and compared to that of other available WGS sequence data from ocean water at various depths. Overall, they found transposon density increases with ocean depth and proposed a relaxation of purifying selection at deeper ocean depths allows the proliferation of these ‘selfish’ gene elements [51]. In other words, the transposons are simply unchecked by negative (purifying) selection. Although they are viewed as deleterious, they are allowed to grow in number due to the lack of competition between organisms. However, nutrients are scarce in the deep ocean and energy is at a premium: in such situations, organisms tend toward more efficient genomes rather than allow them to become disordered. In fact, an explanation based on the assumed role of purifying selection highlights another interesting and important problem in marine microbial ecology: what is the source of genetic diversity in microbial populations? Naively, one would expect that in the narrowly-defined environmental niche of the ocean where there is a limited supply of nutrient and light, the number of coexisting species, in equilibrium, cannot be greater than the number of limiting constraints [45].

Observations are in sharp disagreement with this selection-based idea, and the contradiction has been a vigorous source of debate in the biological literature. Amongst possible proposed solutions to the paradox are spatial and temporal environmental variability [71] and phage predation [68, 69], but a satisfying quantitative explanation is still lacking.

We propose here an alternative explanation for the apparent increase in transposons with ocean depth, one tied to the notion of evolutionary temperature. HGT provides a means of rapid evolution, both increasing the overall mutation rate and transferring functional genetic material between organisms. Examples of viruses encoding important functional genes in the ocean include cyanophage [77] and the photosystem II core reaction [54]. As prevalent as this may seem in the Epipelagic zone of the ocean, microbial population densities decrease inversely with ocean depth soon thereafter [51]. With reduced average density, the opportunities for HGT also decrease. On the other hand, transposons shuffle genetic material within a cell, and this serves to enhance organismal adaptability. One might anticipate, therefore, that to maintain a collection of interacting genomes at the same temperature, or equivalently, at the same level of disorder, contributions from all varieties of MGE need to be included. In particular, as the HGT rate goes down, the density of other MGEs, principally transposons would be expected to increase to compensate.

This argument can be translated into a rough scaling argument, as follows. The number density  $\rho$  of cells is a decreasing function of depth, falling off roughly as  $\rho \sim d^{-1}$  where  $d$  is the depth in the ocean, according to the data [51]. The probability of HGT events per base pair, either conjugal or mediated by an intermediary,  $P_{HGT} \sim \rho^2$ , assuming some sort of law of mass action. On the other hand, transposons shuffle genes within a cell's genome, and this process actually requires two IS elements in order for the transposition event to take place along the genome, so that the probability of each transposition event is proportional to the square of the number density of IS elements,  $\rho_{IS}$ . Making the assumption that there is a uniform genome entropy with depth, and that the HGT and transposition events are independent, we obtain that

$$P_{HGT} \times \rho_{IS}^2 \sim \text{constant} < 1. \quad (1)$$

Making the usual sort of mean field approximation, we then obtain that

$$\rho^2 \times \rho_{IS}^2 \sim \text{constant} \quad (2)$$

and thus that  $\rho_{IS} \sim d$ , a result in rough agreement with the available data.

In order to test this idea, we simulated populations of microbes under continuous selection pressure for a fixed target sequence at different population densities. A schematic of the processes that impact our simulations is given by Fig. 1. For initial conditions, we take genomes of length 300 whose letters are selected randomly from an alphabet size of 20. These genomes are each initialized randomly and a variable number  $N$  of them are placed into an array of size 10000. The population in each subset then remains fixed, but competition is allowed whereby a fitter organism may replace a less fit organism. Point mutation, deletion, HGT, and transposons are allowed. Point mutation randomly exchanges one letter of the genome for another randomly assigned letter while deletion removes the letter entirely. HGT randomly selects a segment 10 letters long from one genome and inserts it into another. The model for transposon behavior is based on Insertion Sequence (IS) element behavior. In this model, an IS is represented by a specific 2 letter combination. As shown in Fig. 2, an IS can non-conservatively insert (i.e., copy-paste as opposed to cut-paste) itself elsewhere within a genome. When 2 IS elements are within a fixed distance of each other (20 in this simulation), then they either transpose the entire length of genome in between including



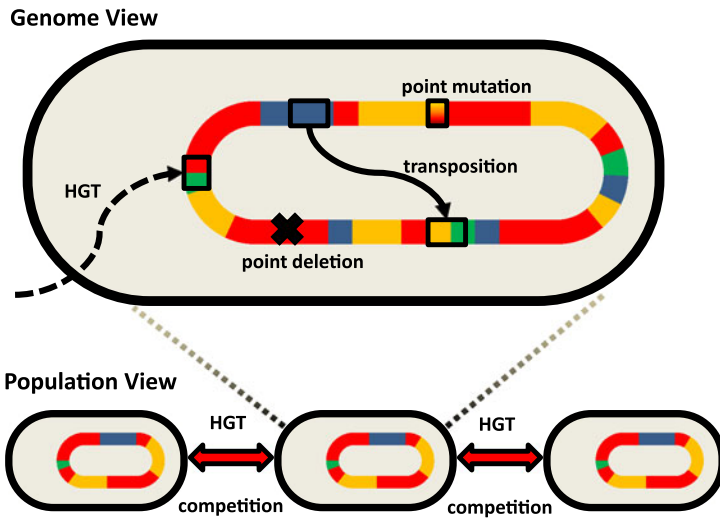


Fig. 1 Schematic of processes present in simulation

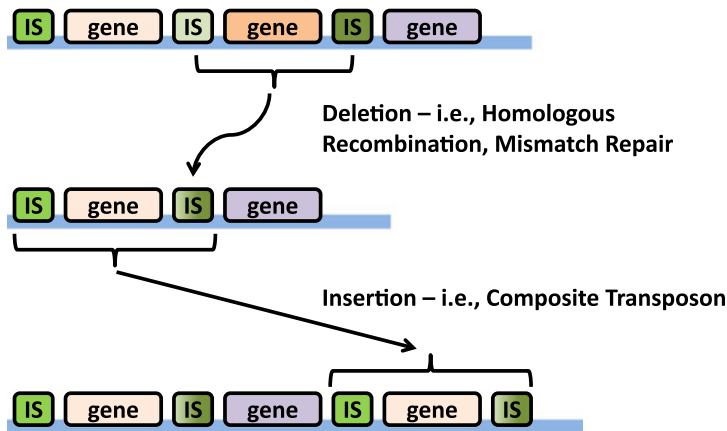


Fig. 2 Schematic of how insertion sequences (IS) act to transpose and delete genetic material

themselves or the region between them is deleted through homologous recombination, each event occurring with equal probability. The rates of mutation, deletion, and transposition are then fixed at 0.001, 3.0, and 0.5 per generation per organism. The rate of HGT is fixed at 1.0 per opportunity. In order for HGT or competition (whereby one organism overwrites another) to occur, two slots in the population array are selected randomly. If both slots contain a living organism, then HGT occurs according to the HGT rate. In this way, as population decrease, so does the number of HGT opportunities. The parameters were chosen with an eye toward obtaining the correct relative rates for each of the processes, as mimicking the orders of magnitude difference between mutation rate and HGT ( $10^9$ ) becomes impractical computationally. We examined mutation rate and system size scaling in for a related system and found that our results were qualitatively robust [15].

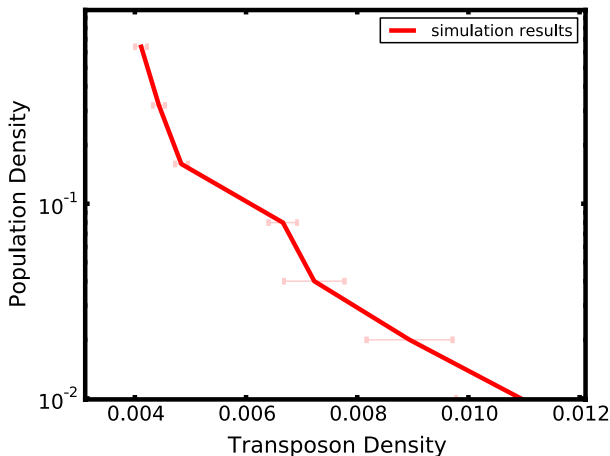
Genes within the genome are identified as a gapless subset of the genome that shares a common subsequence with the target sequence, which is of length 10. The organismal fitness is then given by

$$F = \min\left(\sum_{i=0}^n g_i, 1\right) - n\mu \quad (3)$$

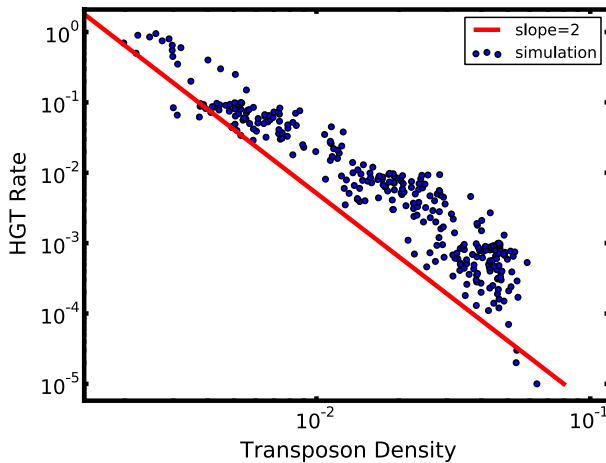
where  $g_i$  is the length of the  $i$ th gene (note that, here, length relates directly to the number of consecutive letters that match the target sequence),  $n$  is the total number of genes within a genome, and  $\mu$  is a penalty of 0.2 for each gene copy present in the genome. This fitness rewards genomes containing subsequences matching a particular target sequence. To some extent, there can be a fitness gain from multiple copies of a gene, but this gain is not limitless and many short matches to the target sequence are penalized. The fitness function described here has a basis in a recently proposed model for the evolution of novel gene function [11]. A similar fitness function was also at the basis of related theoretical work on the transposon dynamics of recent obligate associations [15].

Figure 3 plots the results from our simulation of microbial competition according to the rules outlined above across a population density gradient. This tests the hypothesis that the apparent increase in transposon density is related to decreasing levels of HGT due to the decreasing population densities in deeper waters. As shown, we do indeed see an increase in the transposon density that corresponds with decreasing population density. However, in order to assert that this is due to the lack of HGT and not other effects such as population size or competition timescales (which have also slowed down due to competition requiring two organisms to interact), we must isolate these other effects and focus on HGT alone.

Figure 4 shows what happens when we allow a fixed population size to adapt under different HGT rates. The trend of increasing transposon density holds for decreasing HGT



**Fig. 3** Transposon density versus population density. We simulated the adaptation of organisms toward a fixed target sequence allowing point mutation, deletion, HGT, and transposon dynamics for 20,000 generations and plot the averaged final transposon densities of 100 simulations for each point. Different population densities have differing amounts of opportunities for HGT. Our simulation results show that at lower population densities with decreased HGT transposons increase in relative abundance. This is consistent with the idea that transposons serve as a substitute for the evolutionary dynamic provided by HGT in shallower waters. In ocean waters, population density is inversely proportional to depth below the Epipelagic Zone (>200 m), to first order approximation [51]



**Fig. 4** Transposon density versus HGT rate for fixed population size. Keeping population size at the fixed value of 1,000, we plot the transposon density after 10,000 generations averaged across 100 simulations per point. The deletion rate is 2 per generation per organism. All other parameters are the same as described in the main text. In order to verify that the increase in transposon density seen in Fig. 3 is the result of an HGT:transposon evolutionary tradeoff, we eliminate other sources of variation by fixing the population size and varying the HGT rate. This serves to eliminate the effects of differing population sizes and competition timescales present in the previous simulation. In this simulation, we show increases in transposon density with decreasing HGT rate. We plot a power law *line* with an exponent of 2 for reference. The *line* does not represent a fit to the data

rate. This is consistent with our hypothesis that the transposon density trend seen in the ocean corresponds to lower HGT rates in the ocean depths. Reduction of HGT leads to increased transposon numbers as originally postulated.

The possibility that some notion of ‘adaptability’ underlies transposon dynamics points toward two very interesting ideas. The first is that transposons, as well as other mobile genetic elements such as viruses and plasmids, could be more than just parasitic gene sequences feeding off host genomes. Instead, they may be required for the long term survival, adaptability, and diversification of an organismal lineage. The second is that evolutionary dynamics such as transposon proliferation may be driven by generic processes rather than governed by the specific histories of individual populations. This means that the properties of biological organism can and should be understood from the viewpoint of the statistics of a physical process rather than the particulars of a historical accident.

## 6 Summary

HGT couples together unrelated organismal lineages in a way that requires us to rethink the classical point-mutation based notions of population genetics. HGT has been shown to be a prevalent force in microbial evolution especially. The impact of HGT on gene and organismal evolution has not been fully understood. However, notions from statistical physics such as temperature and annealing have been applied to evolutionary dynamics before and will no doubt continue to play a role in determining the underlying rules of evolution. We showed one example of how theory can play a role by offering up a quantitative hypothesis on the role of HGT in determining how transposon densities vary with ocean depth. Our example

involves very generic interactions and does not depend on the particular environment or microbial species. It is exactly these non-specific types of studies that are important if we are to unify our understanding of the dynamics of genome evolution.

**Acknowledgements** We thank Ed DeLong, Carl Woese and Nicholas Guttenberg for valuable discussions, and Pan-Jun Kim and Zhenyu Wang for helpful comments on the manuscript. This work was partially supported by the National Science Foundation through grant NSF-EF-0526747.

## References

- Adami, C.: Information theory in molecular biology. *Phys. Life Rev.* **1**(1), 3–22 (2004)
- Adami, C., Ofria, C., Collier, T.: Evolution of biological complexity. *Proc. Natl. Acad. Sci. USA* **97**(9), 4463–4468 (2000)
- Aminov, R.I., Mackie, R.I.: Evolution and ecology of antibiotic resistance genes. *FEMS Microbiol. Lett.* **271**(2), 147–161 (2007)
- Anderson, E.S.: Possible importance of transfer factors in bacterial evolution. *Nature* **209**, 637–638 (1966)
- Anderson, N.G.: Evolutionary significance of virus infection. *Nature* **227**, 1346–1347 (1970)
- Babic, A., Lindner, A.B., Vulic, M., Stewart, E.J., Radman, M.: Direct visualization of horizontal gene transfer. *Science* **319**(5869), 1533–1536 (2008)
- Beiko, R.G., Harlow, T.J., Ragan, M.A.: Highways of gene sharing in prokaryotes. *Proc. Natl. Acad. Sci. USA* **102**(40), 14332–14338 (2005)
- Beltrami, E., Carroll, T.: Modeling the role of viral disease in recurrent phytoplankton blooms. *J. Math. Biol.* **32**(8), 857–863 (1994)
- Beman, J.M., Arrigo, K.R., Matson, P.A.: Agricultural runoff fuels large phytoplankton blooms in vulnerable areas of the ocean. *Nature* **434**(7030), 211–214 (2005)
- Bennett, P.M.: Plasmid encoded antibiotic resistance: acquisition and transfer of antibiotic resistance genes in bacteria. *Br. J. Pharmacol.* **153**(S1), S347–S357 (2008)
- Berghorsson, U., Andersson, D.I., Roth, J.R.: Ohno's dilemma: Evolution of new genes under continuous selection. *Proc. Natl. Acad. Sci. USA* **104**(43), 17004 (2007)
- Brochier, C., Forterre, P., Gribaldo, S.: An emerging phylogenetic core of archaea: phylogenies of transcription and translation machineries converge following addition of new genome sequences. *BMC Evol. Biol.* **5**(1), 36 (2005)
- Butler, T., Goldenfeld, N.: Robust ecological pattern formation induced by demographic noise. *Phys. Rev. E* **80**(3), 30902 (2009)
- Chen, I., Dubnau, D.: DNA uptake during bacterial transformation. *Nat. Rev., Microbiol.* **2**(3), 241–249 (2004)
- Chia, N., Goldenfeld, N.: The dynamics of gene duplication and transposons in microbial genomes following a sudden environmental change. [arXiv:1005.3349](https://arxiv.org/abs/1005.3349) (2010)
- Chia, N., Woese, C.R., Goldenfeld, N.: A collective mechanism for phase variation in biofilms. *Proc. Natl. Acad. Sci. USA* **105**(38), 14597–14603 (2008)
- Chia, N., Cann, I., Olsen, G.J.: Evolution of dna replication protein complexes in eukaryotes and archaea. *PLoS ONE* **5**(6), e10866 (2010)
- Cohen, O., Pupko, T.: Inference and characterization of horizontally transferred gene families using stochastic mapping. *Mol. Biol. Evol.* **27**(3), 703 (2010)
- Davis, J.J., Olsen, G.J.: Modal codon usage: Assessing the typical codon usage of a genome. *Mol. Biol. Evol.* **27**(4), 800 (2010)
- De La Torre, J.R., Christianson, L.M., Béjà, O., Suzuki, M.T., Karl, D.M., Heidelberg, J., DeLong, E.F.: Proterorhodopsin genes are distributed among divergent marine bacterial taxa. *Proc. Natl. Acad. Sci. USA* **100**(22), 12830 (2003)
- DeLong, E.: Microbial community genomics in the ocean. *Nat. Rev., Microbiol.* **3**(6), 459–469 (2005)
- Denamur, E., Matic, I.: Evolution of mutation rates in bacteria. *Mol. Microbiol.* **60**(4), 820–827 (2006)
- Dewar, R.C., Porte, A.: Statistical mechanics unifies different ecological patterns. *J. Theor. Biol.* **251**(3), 389–403 (2008)
- Edwards, R., Rohwer, F.: Viral metagenomics. *Nat. Rev., Microbiol.* **3**(6), 504–510 (2005)
- Eisen, J.A.: Horizontal gene transfer among microbial genomes: new insights from complete genome analysis. *Curr. Opin. Genet. Dev.* **10**(6), 606–611 (2000)
- Elsas, J.D., Bailey, M.J.: The ecology of transfer of mobile genetic elements. *FEMS Microbiol. Ecol.* **42**(2), 187–197 (2002)

27. Farahi, K., Pusch, G.D., Overbeek, R., Whitman, W.B.: Detection of lateral gene transfer events in the prokaryotic trna synthetases by the ratios of evolutionary distances method. *J. Mol. Evol.* **58**(5), 615–631 (2004)
28. Frigaard, N.U., Martinez, A., Mincer, T.J., DeLong, E.F.: Proteorhodopsin lateral gene transfer between marine planktonic Bacteria and Archaea. *Nature* **439**(7078), 847–850 (2006)
29. Frost, L.S., Leplae, R., Summers, A.O., Toussaint, A.: Mobile genetic elements: the agents of open source evolution. *Nat. Rev., Microbiol.* **3**(9), 722–732 (2005)
30. Garcia-Vallvé, S., Romeu, A., Palau, J.: Horizontal gene transfer in bacterial and archaeal complete genomes. *Genome Res.* **10**(11), 1719 (2000)
31. Gladyshev, E., Meselson, M., Arkipova, I.: Massive horizontal gene transfer in bdelloid rotifers. *Science* **320**(5880), 1210 (2008)
32. Gogarten, J.P., Townsend, J.P.: Horizontal gene transfer genome innovation and evolution. *Nat. Rev., Microbiol.* **3**(9), 679–687 (2005)
33. Goldenfeld, N., Woese, C.: Biology's next revolution. *Nature* **445**(7126), 369 (2007)
34. Goldenfeld, N., Woese, C.: Life is physics: evolution as a collective phenomenon far from equilibrium. *Ann. Rev. Condens. Matter. Phys.* **1** (2010, in press)
35. Hallegraef, G.: A review of harmful algal blooms and their apparent global increase. *Phycologia* **32**(2), 79–99 (1993)
36. Handelsman, J.: Metagenomics: application of genomics to uncultured microorganisms. *Microbiol. Mol. Biol. Rev.* **68**(4), 669–685 (2004)
37. Hartwell, L., Hopfield, J., Leibler, S., Murray, A.: From molecular to modular cell biology. *Nature* **402**, C47–C52 (1999)
38. He, J., Sun, J., Deem, M.: Spontaneous emergence of modularity in a model of evolving individuals and in real networks. *Phys. Rev. E, Stat. Nonlinear Soft Matter Phys.* **79**(3), 031907 (2009)
39. Hecht, I., Ben-Jacob, E., Levine, H.: Correlated phenotypic transitions to competence in bacterial colonies. *Phys. Rev. E* **76**(4), 40901 (2007)
40. Hendrix, R.W., Lawrence, J.G., Hatfull, G.F., Casjens, S.: The origins and ongoing evolution of viruses. *Trends Microbiol.* **8**(11), 504–508 (2000)
41. Holden, M.T.G., Feil, E.J., Lindsay, J.A., Peacock, S.J., Day, N.P.J., Enright, M.C., Foster, T.J., Moore, C.E., Hurst, L., Atkin, R., et al.: Complete genomes of two clinical staphylococcus aureus strains: evidence for the rapid evolution of virulence and drug resistance. *Proc. Natl. Acad. Sci. USA* **101**(26), 9786–9792 (2004)
42. Hotopp, J., Clark, M., Oliveira, D., Foster, J., Fischer, P., Torres, M., Giebel, J., Kumar, N., Ishmael, N., Wang, S., et al.: Widespread lateral gene transfer from intracellular bacteria to multicellular eukaryotes. *Science* **317**(5845), 1753–1756 (2007)
43. Hubbell, S.P.: *The Unified Neutral Theory of Species Abundance and Diversity*. Princeton University, Princeton (2001)
44. Huppert, A., Blasius, B., Stone, L.: A model of phytoplankton blooms. *Am. Nat.* **159**(2), 156–171 (2002)
45. Hutchinson, G.: The paradox of the plankton. *Am. Nat.* **95**(882), 137–145 (1961)
46. Jeong, H., Tombor, B., Albert, R., Oltvai, Z.N., Barabási, A.L.: The large-scale organization of metabolic networks. *Nature* **407**(6804), 651–654 (2000)
47. Kashtan, N., Alon, U.: Spontaneous evolution of modularity and network motifs. *Proc. Natl. Acad. Sci. USA* **102**(39), 13773–13778 (2005)
48. Kashtan, N., Parter, M., Dekel, E., Mayo, A., Alon, U.: Extinctions in heterogeneous environments and the evolution of modularity. *Evolution* **63**(8), 1964–1975 (2009)
49. Keeling, P.J., Palmer, J.D.: Horizontal gene transfer in eukaryotic evolution. *Nat. Rev. Genet.* **9**(8), 605–618 (2008)
50. Koike, S., Krapac, I., Oliver, H., Yannarell, A., Chee-Sanford, J., Aminov, R., Mackie, R.: Monitoring and source tracking of tetracycline resistance genes in lagoons and groundwater adjacent to swine production facilities over a 3-year period. *Appl. Environ. Microbiol.* **73**(15), 4813–4823 (2007)
51. Konstantinidis, K.T., Bruff, J., Karl, D.M., DeLong, E.F.: Comparative metagenomic analysis of a microbial community residing at a depth of 4,000 meters at station ALOHA in the North Pacific subtropical gyre. *Appl. Environ. Microbiol.* **75**(16), 5345–5355 (2009)
52. Kreimer, A., Borenstein, E., Gophna, U., Ruppin, E.: The evolution of modularity in bacterial metabolic networks. *Proc. Natl. Acad. Sci. USA* **105**(19), 6976–6981 (2008)
53. Kunin, V., Goldovsky, L., Darzentas, N., Ouzounis, C.A.: The net of life: reconstructing the microbial phylogenetic network. *Genome Res.* **15**(7), 954–959 (2005)
54. Lindell, D., Sullivan, M.B., Johnson, Z.I., Tolonen, A.C., Rohwer, F., Chisholm, S.W.: Transfer of photosynthesis genes to and from *Prochlorococcus* viruses. *Proc. Natl. Acad. Sci. USA* **101**(30), 11013–11019 (2004)

55. Lorenz, M., Wackernagel, W.: Bacterial gene transfer by natural genetic transformation in the environment. *Microbiol. Mol. Biol. Rev.* **58**(3), 563 (1994)
56. Lorenz, D.M., Jeng, A., Deem, M.W.: The emergence of modularity in biological systems. *J. Life Rev.* (2010, submitted)
57. Mahillon, J., Chandler, M.: Insertion sequences. *Microbiol. Mol. Biol. Rev.* **62**(3), 725 (1998)
58. McKenzie, G.J., Harris, R.S., Lee, P.L., Rosenberg, S.M.: The *sos* response regulates adaptive mutation. *Proc. Natl. Acad. Sci. USA* **97**(12), 6646 (2000)
59. Médigue, C., Rouxel, T., Vigier, P., Hénaut, A., Danchin, A.: Evidence for horizontal gene transfer in *Escherichia coli* speciation. *J. Mol. Biol.* **222**(4), 851–856 (1991)
60. Menge, D.N.L., Weitz, J.S.: Dangerous nutrients: Evolution of phytoplankton resource uptake subject to virus attack. *J. Theor. Biol.* **257**(1), 104–115 (2009)
61. Mirollo, R.E., Strogatz, S.H.: Synchronization of pulse-coupled biological oscillators. *SIAM J. Appl. Math.* **50**(6), 1645–1662 (1990)
62. Monier, A., Pagarete, A., de Vargas, C., Allen, M., Read, B., Claverie, J., Ogata, H.: Horizontal gene transfer of an entire metabolic pathway between a eukaryotic alga and its DNA virus. *Genome Res.* **19**(8), 1441 (2009)
63. Nielsen, K.M., Townsend, J.P.: Monitoring and modeling horizontal gene transfer. *Nat. Biotechnol.* **22**(9), 1110–1114 (2004)
64. Pace, J., Gilbert, C., Clark, M., Feschotte, C.: Repeated horizontal transfer of a DNA transposon in mammals and other tetrapods. *Proc. Natl. Acad. Sci. USA* **105**(44), 17023 (2008)
65. Pál, C., Papp, B., Lercher, M.: Adaptive evolution of bacterial metabolic networks by horizontal gene transfer. *Nat. Genet.* **37**(12), 1372–1375 (2005)
66. Parter, M., Kashtan, N., Alon, U.: Environmental variability and modularity of bacterial metabolic networks. *BMC Evol. Biol.* **7**, 169/1–8 (2007)
67. Prangishvili, D., Forterre, P., Garrett, R.A.: Viruses of the archaea: a unifying view. *Nat. Rev., Microbiol.* **4**(11), 837–848 (2006)
68. Rodriguez-Brito, B., Li, L., Wegley, L., Furlan, M., Angly, F., Breitbart, M., Buchanan, J., Desnues, C., Dinsdale, E., Edwards, R., Felts, B., Haynes, M., Liu, H., Lipson, D., Mahaffy, J., Belen Martin-Cuadrado, A., Mira, A., Nulton, J., et al.: Viral and microbial community dynamics in four aquatic environments. *ISME J.* **4**(6), 739–751 (2010)
69. Rodriguez-Valera, F., Martin-Cuadrado, A., Rodriguez-Brito, B., Pasic, L., Thingstad, T., Rohwer, F., Mira, A.: Explaining microbial population genomics through phage predation. *Nat. Rev., Microbiol.* **7**(11), 828–836 (2009)
70. Salyers, A.A., Amabile-Cuevas, C.F.: Why are antibiotic resistance genes so resistant to elimination? *Antimicrob. Agents Chemother.* **41**(11), 2321 (1997)
71. Scheffer, M., Rinaldi, S., Huisman, J., Weissing, F.: Why plankton communities have no equilibrium: solutions to the paradox. *Hydrobiologia* **491**(1), 9–18 (2003)
72. Schoemann, V., Becquevort, S., Stefels, J., Rousseau, V., Lancelot, C.: *Phaeocystis* blooms in the global ocean and their controlling mechanisms: a review. *J. Sea Res.* **53**(1–2), 43–66 (2005)
73. Simon, H.: The architecture of complexity. *Proc. Am. Philos. Soc.* **106**(6), 467–482 (1962)
74. Snel, B., Bork, P., Huynen, M.A.: Genomes in flux: the evolution of archaeal and proteobacterial gene content. *Genome Res.* **12**(1), 17–25 (2002)
75. Sonea, S.: A bacterial way of life. *Nature* **331**(6153), 216 (1988)
76. Sørensen, S.J., Bailey, M., Hansen, L.H., Kroer, N., Wuertz, S.: Studying plasmid horizontal transfer in situ: a critical review. *Nat. Rev., Microbiol.* **3**(9), 700–710 (2005)
77. Sullivan, M.B., Waterbury, J.B., Chisholm, S.W.: Cyanophages infecting the oceanic cyanobacterium *Prochlorococcus*. *Nature* **424**(6952), 1047–1051 (2003)
78. Sun, J., Deem, M.W.: Spontaneous emergence of modularity in a model of evolving individuals. *Phys. Rev. Lett.* **99**(22), 228107 (2007)
79. Suttle, C.A.: Viruses in the sea. *Nature* **437**(7057), 356–361 (2005)
80. Suttle, C.A.: Marine viruses—major players in the global ecosystem. *Nat. Rev., Microbiol.* **5**(10), 801–812 (2007)
81. Syvanen, M.: Horizontal gene transfer: evidence and possible consequences. *Annu. Rev. Genet.* **28**(1), 237–261 (1994)
82. Thomas, C.M., Nielsen, K.M.: Mechanisms of, and barriers to, horizontal gene transfer between bacteria. *Nat. Rev., Microbiol.* **3**(9), 711–721 (2005)
83. Thomas, R., Thieffry, D., Kaufman, M.: Dynamical behaviour of biological regulatory networks. I. Biological role of feedback loops and practical use of the concept of the loop-characteristic state. *Bull. Math. Biol.* **57**(2), 247–276 (1995)
84. Tsigos, A., Rigoutsos, I.: A new computational method for the detection of horizontal gene transfer events. *Nucleic Acids Res.* **33**(3), 922 (2005)

85. Tyson, G., Chapman, J., Hugenholtz, P., Allen, E., Ram, R., Richardson, P., Solovyev, V., Rubin, E., Rokhsar, D., Banfield, J.: Community structure and metabolism through reconstruction of microbial genomes from the environment. *Nature* **428**(6978), 37–43 (2004)
86. Vetsigian, K., Goldenfeld, N.: Global divergence of microbial genome sequences mediated by propagating fronts. *Proc. Natl. Acad. Sci. USA* **102**(20), 7332 (2005)
87. Vetsigian, K., Woese, C., Goldenfeld, N.: Collective evolution and the genetic code. *Proc. Natl. Acad. Sci. USA* **103**(28), 10696–10702 (2006)
88. Wagner, G., Pavlicev, M., Cheverud, J.: The road to modularity. *Nat. Rev. Genet.* **8**(12), 921–931 (2007)
89. Waters, C., Bassler, B.: Quorum sensing: cell-to-cell communication in bacteria. *Annu. Rev. Cell Dev. Biol.* **21**, 319–46 (2005)
90. Weinbauer, M.G.: Ecology of prokaryotic viruses. *FEMS Microbiol. Rev.* **28**(2), 127–181 (2004)
91. Weinbauer, M.G., Rassoulzadegan, F.: Are viruses driving microbial diversification and diversity? *Environ. Microbiol.* **6**(1), 1–11 (2004)
92. Wilmes, P., Simmons, S.L., Deneff, V.J., Banfield, J.F.: The dynamic genetic repertoire of microbial communities. *FEMS Microbiol. Rev.* **33**(1), 109–132 (2009)
93. Woese, C.R.: On the evolution of cells. *Proc. Natl. Acad. Sci. USA* **99**(13), 8742–8748 (2002)
94. Woese, C.R., Fox, G.E.: The concept of cellular evolution. *J. Mol. Evol.* **10**(1), 1–6 (1977)
95. Woese, C.R., Fox, G.E.: Phylogenetic structure of the prokaryotic domain: the primary kingdoms. *Proc. Natl. Acad. Sci. USA* **74**(11), 5088 (1977)
96. Woese, C.R., Olsen, G.J., Ibba, M., Soll, D.: Aminoacyl-tRNA synthetases, the genetic code, and the evolutionary process. *Microbiol. Mol. Biol. Rev.* **64**(1), 202 (2000)